

1TM8 =>superSeed =>1YMG 2B5F**Fig. 1. Tetramer and monomer structure of bAQP0.**

(Upper Left) Cartoon of the bAQP0 tetramer looking down the zaxis from the extracellular side of the protein. Yellow and blue indicate structures derived from each of the two gene-duplicated portions of the primary sequence. (Upper Right) Cartoon showing the same view as in Upper Left, with each monomer shown in a different representation. (Lower) Cartoon of an bAQP0 monomer in a side view, with the uppermost extracellular side in crossed eye stereo. All images were made with pymol(DeLano Scientific, San Carlos, CA)

The structure starts with residue 6 and ends with residue 239, representing 88% of the molecule. Because the complete primary sequence of b AQP0 is intact as determined by MS, five residues at the N terminus and 24 residues at

the C terminus are not well ordered in the structure. The useful electron density ends at residue 239, and the N and C termini lie on the cytoplasmic side of the membrane.

The 28-Å-long, cylindrical bAQP0 channel is flanked by shallow vestibules on each end (Fig. 2). Starting from the extracellular side, the vestibule narrows to a diameter of <math><10 \text{ \AA}</math> between residues Asn-115, Thr-120, and His-34. His-34 is oriented into the center line of the channel and is responsible for most of the narrowing of the vestibule. The channel narrows a diameter of 1.99 Å at residues Phe-48(Trp-48), His-172(Ile-190), Met-176(Gly-195), Ala-181(Phe-200), and Arg-187(206). In AQP1, AQPZ, and GlpF, this region is the narrowest region of the channel (Fig. 3) (9, 22, 23). four backbone carbonyls of successive residues [Gly-180(Thr-198), Ala-181(Gly-199), Gly-182(Phe-200), and Met-183(Ala-201)] provide the canonical AQP Hydrogen bond acceptors that align waters through the channel. The OD1 of Asn-119(Val-129), NH₂ of Arg-187(Ala-205), and His-172(Ile-190) provide donor Hydrogen bonds for the waters. These residues bind four ordered water molecules (Fig. 2 Center) and further orient the channel waters. Farther into the channel, the side chain of Tyr-23(Leu-21) is oriented directly toward the central axis of the channel and, with Phe-141(Leu-159), Leu-52(Val-52), and Leu-168(186), constricts the channel diameter to 2.5 Å. Just after the Tyr-23 constriction are Asn-68(68) and Asn-184(203) of the signature NPA–NPA motifs that orient the key central water molecule that is responsible for preventing the reorientation that would be necessary for any proton conduction. On the cytoplasmic side of the NPAs, the line of backbone carbonyl oxygens resumes along one wall of the channel from Gly-64(64), Ala-65(65), His-66(66), and Gly-67(67), and it ends at Tyr-149(Thr-167). The line of eight backbone carbonyls and other channel-lining residues establishes a tight single-file pathway for water highlighted by the eight water molecules in the channel.

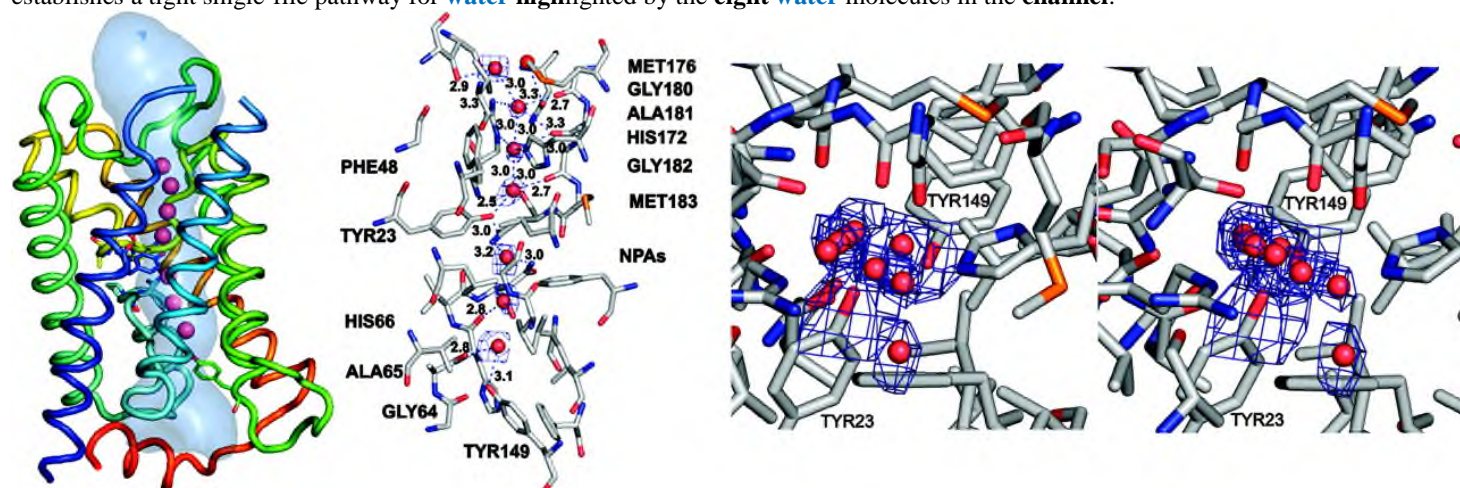


Fig. 2. Monomer channel views of bAQP0. (Left) Side view of the monomer and water molecules (red spheres) in the channel. The channel luminal surface is shown in light blue. Each helix is colored in order of the rainbow. (Center) Side view looking from the midmembrane plane toward the monomer channel residues and water molecules in the channel. Helix bonds to the channel waters are shown as dotted lines. Electron density around waters is shown in a composite omit 2Fo–Fc map contoured at 0.5 σ for clarity. (Right) Stereo view from the extracellular side of the channel. (Electron density around waters and Tyr-23 and Tyr-149 are contoured at 0.5 σ . All images were made with pymol.

Tyr-149(Thr-149) points directly into the channel and, together with Val-56(Ile-56), Gly-64(64), His-66(66), and

Phe-75(Leu-75), forms another **constriction** that is the **narrowest region** of the **channel**. It accepts a sphere with a maximum diameter of 1.5 Å (as determined by using the program hole; ref. 21). In **AQP0**, it serves as a **cytoplasmic end** of the **narrow** part of the **channel**. Continuing in toward the **cytoplasmic side**, the **channel** widens slightly to accept a sphere with an average diameter of 3 Å, which is significantly **narrower** than in other **AQP** structures (**AQP1**, 3.5–4.0 Å; **GlpF**, 4.0–5.0 Å) (22, 23). The **two residues Tyr-23(Leu-21)** and **Tyr-149(Thr-149)** are in quasi-2-fold related positions evoked by gene duplication, and in the other **AQPs**, **Tyr-23(Leu-21)** is either a phenylalanine or a leucine and **Tyr-149(Thr-149)** is either a **threonine** or **leucine**.

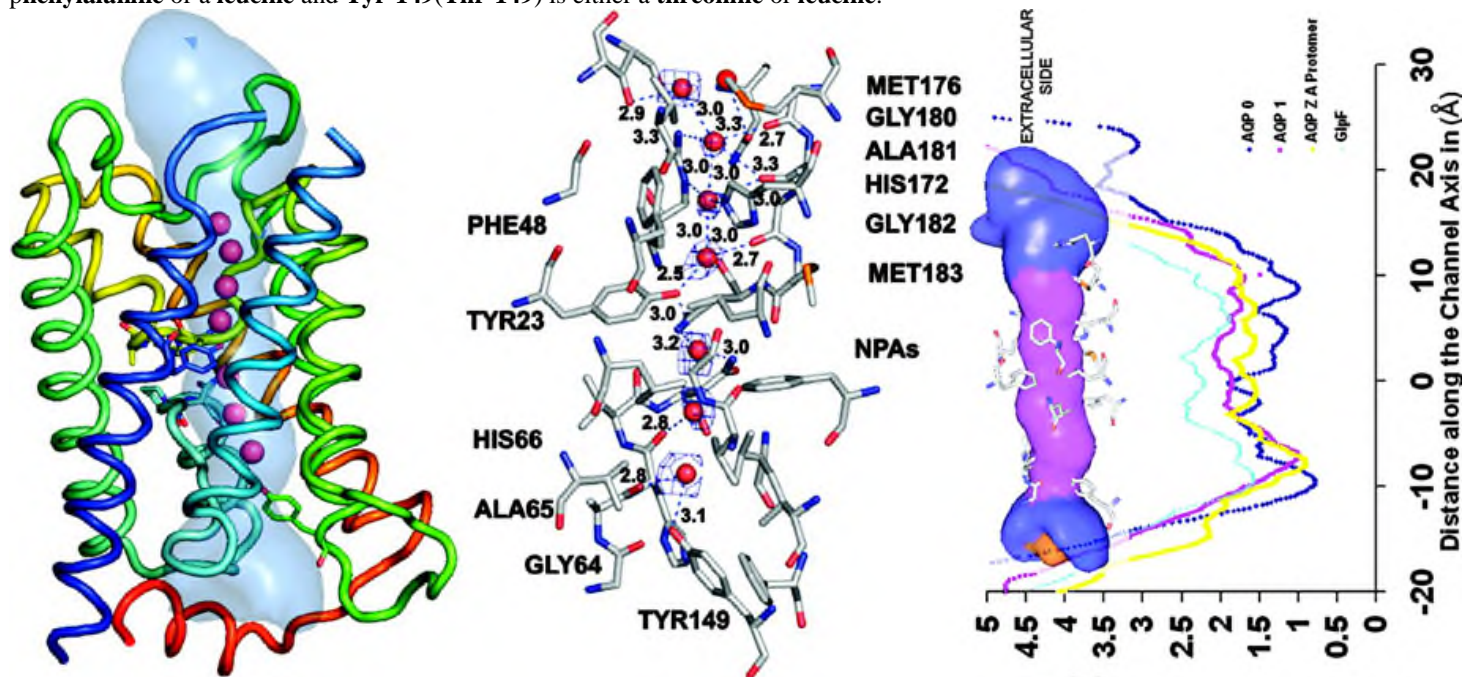


Fig. 3. **Channel radius profile plot.** Channel radius profiles of AQPs of known structure with corresponding structural elements are shown (22, 23). The AQPZ “A” protomer was used for radius calculations for AQPZ. The distance along the **channel** axis is calculated by using a point **midway** between the **Asn-Pro-Ala** sequences (NPAs) as the **zero** point. **Radii** were calculated with hole (39). **Channel** volume is shown in the background, with major **channel-forming residues**. The **pink central region** has a diameter of <2.5 Å, the **blue region** has a diameter of >2.5 Å and <10 Å. All images were made with pymol.

Regulation of Water Conductance by pH, Ions, or Posttranslational Modification? Modulation by changes in **pH** or **ion concentration** have been suggested in the control of **transport** of **water** and possibly other **substrates** **O₂**, **CO₂**, **NO**, **CO** (10,11,13,30–35) through **AQP0**. However, Boron and coworkers (13) argue against these effects and show zero dependence on **pH** or **Ca²⁺**. **His-40** has been suggested to be the residue that is responsible for the inhibitory effect of **pH** or **Ca²⁺** on **water** transport rates (11–13). Therefore, Nemeth-Cahalan and Hall made mutations that **changed His-40** to **alanine**, **aspartate**, or **lysine**, and they showed that treatment of **oocytes** expressing the mutants no longer displayed the **pH-dependent** closing as **pH** was raised to >6.5 . Reaction of **histidines** in **AQP0** with diethylpyrocarbonate (DEPC) removed **pH** dependence and actually increased **water** conductance (restored by **histidine**-specific reversal with **hydroxylamine**), adding support to the case that titration of **histidines** alters **conductance** (11, 12).

The comparison of the **two** structures (**pH 10** vs. **6.0**) brackets the range of observed **pH** dependence and, therefore, should reveal any structural consequences of **His** titration. The comparison shows no positional difference of **His-40** in the **extracellular vestibule**, **His-172** (which lies in the wall of the **channel**), or **His-66** in the **cytoplasmic vestibule** (where it is the **Helix bond** acceptor from **Thr-72**). Because of their **water**-accessible position, one could expect the **pK_a** of **His-40** and **His-66** to be relatively normal and unlikely to be responsible for **gating** at **higher pH** without structural **change** that we do not see at **pH 10** vs. **6**. **His-172** near the **extracellular constriction** region is **Helix bond donor** to the **second water** in the line of **waters** throughout the **channel** (see Fig. 4 Lower Left). Thus, all three **histidines** show no evidence of **pH-dependent conformational change**. Fig. 4. **Comparisons of the x-ray and electron crystal structures.** (Upper Left) Structures overlaid with **x-ray structure** are shown in **purple**, and the **electron-diffraction structure** is shown in **green**. (Upper Right) View down the **monomer channel z** axis from the **extracellular side** showing the positions of **Tyr-23** and **Tyr-149**. (Lower Left) View down the **monomer channel z** axis from the **extracellular side** showing the positions of the three **histidines** that are close to the **channel** and **vestibules**. (Lower Right) Side view. The **extracellular residues** involved in **cell-to-cell contacts** are **highlighted** and labeled Gly114,Arg113,Val112,Ala111,Pro110,Pro109;His40,Leu39,Pro38,Gly37,Ala35,Trp34,Arg33,Leu32, Ser31;Ser126,Val125,Gly124,Pro123,His122,Leu121,Thr120,Asn119,Asn200,Thr199,Phe198,Asn197,Arg196,Thr195,Leu194.